# Beth Israel Lahey Health Beth Israel Deaconess Medical Center HARVARD MEDICAL SCHOOL BROAD INSTITUTE

- 1: Department of Pathology, Beth Israel Deaconess Medical Center, Boston, MA
- 2: Harvard Medical School, Boston, MA
- 3: Broad Institute of MIT and Harvard, Cambridge, MA
- 4: NanoString Technologies, Inc., Seattle, WA

nanoStrinq

5: Department of Medicine, Beth Israel Deaconess Medical Center, MA Contact email: <u>ivlachos@bidmc.harvard.edu</u>

In addition to causing viral pneumonia and acute respiratory distress syndrome, COVID-19 impacts multiple organ systems, and manifests in heart and renal failure, liver injury, bone marrow suppression, nerve damage, and thromboembolic events. How the virus manifests in various organs remains poorly understood. To this end, we combined in situ tissue transcriptome profiling aided with single nucleus sequencing (sNuc-Seq) to generate integrated atlases on five tissues from five patients who succumbed to COVID-19 at a tertiary medical center. The integrative analyses revealed localized transcriptional programs, capture intercellular communication, and provide novel insights in disease pathogenesis.

Whole transcriptome spatial profiling and sNuc-Seq was performed on lung, heart, liver, kidney, and spleen, from five patients with PCRconfirmed COVID-19 pneumonia. Postmortem biopsies were collected within 3h of asystole under ultrasound guidance. Biopsies were formalin fixed and flash frozen for digital spatial profiling (DSP) and sNuc-Seq, respectively. Whole Transcriptome Atlas (WTA) DSP was performed on FFPE tissue sections using the NanoString GeoMx® platform. sNuc-Seq libraries were prepared on a 10x Chromium Controller using isolated nuclei from frozen tissue. SARS-CoV-2 abundance was quantified with qPCR, probes in WTA library, and In Situ Hybridization.

On average, 5,000+ nuclei were sequenced per sample with >1,200 genes expressed per cell, showcasing the robustness of the sNuc-Seq protocol for diverse tissues and frozen samples. In the spatial analysis, 288 ROIs were quantified across 15 patient samples, providing an average of ~16,000 genes expressed per ROI. sNuc-Seq data permitted the characterization of cellular populations as well as the identification of cell-specific markers and pathways. Importantly, parenchymal cells were extensively represented across tissues, showcasing an additional advantage of the sNuc-Seq protocol. The WTA datasets enabled direct comparisons between microscopic regions with distinct pathological features (e.g. lung alveoli with inflammation vs normal appearing alveoli) and revealed diseasespecific pathways. Spatial transcriptomics also supported the generation of atlases capturing the role of tissue architecture on gene expression. For instance, expression profiles defined by liver zonation were identified by WTA-DSP and directed the annotation of the sNuc-Seg cell clusters, a process which relied in the past mostly on rodent data or a limited set of marker genes. These two modalities exhibited very high complementarity, proving that coupling DSP profiling with single-nucleus assays provides a unique opportunity to understand spatial biological dynamics at a cellular resolution.

COVID-19 exhibits multi-organ phenotypes differentially affecting tissues and cellular programs. The integration of WTA, a novel whole transcriptome spatial profiling assay, with sNuc-Seq maximized the utility of both modalities and mitigated their respective limitations. This integrative approach is broadly applicable and allows mechanistic investigations of complex diseases including COVID-19.

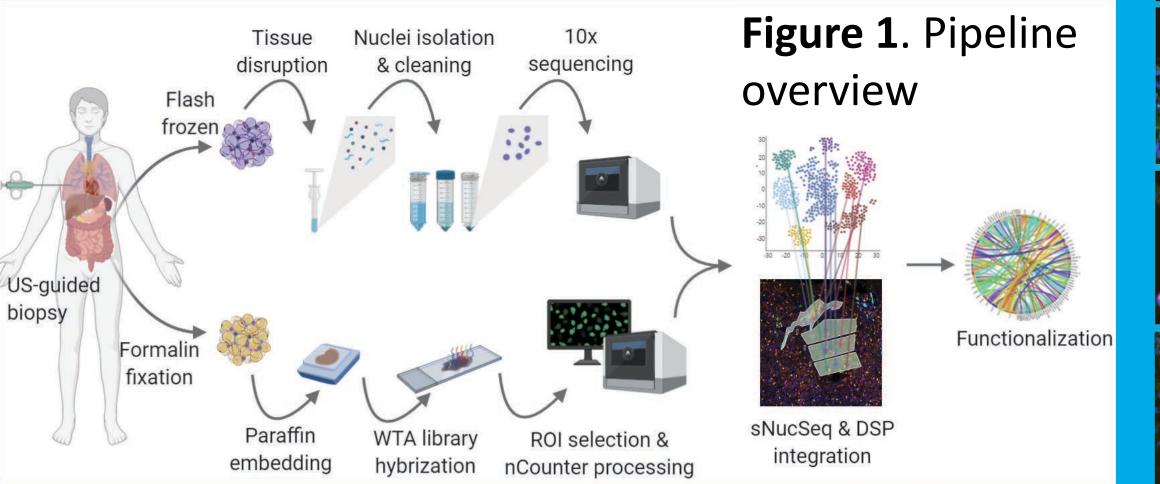
## Integration of Whole Transcriptome Spatial Profiling with Single Nucleus Sequencing reveals unique pathways associated with multi-organ phenotypes in COVID-19

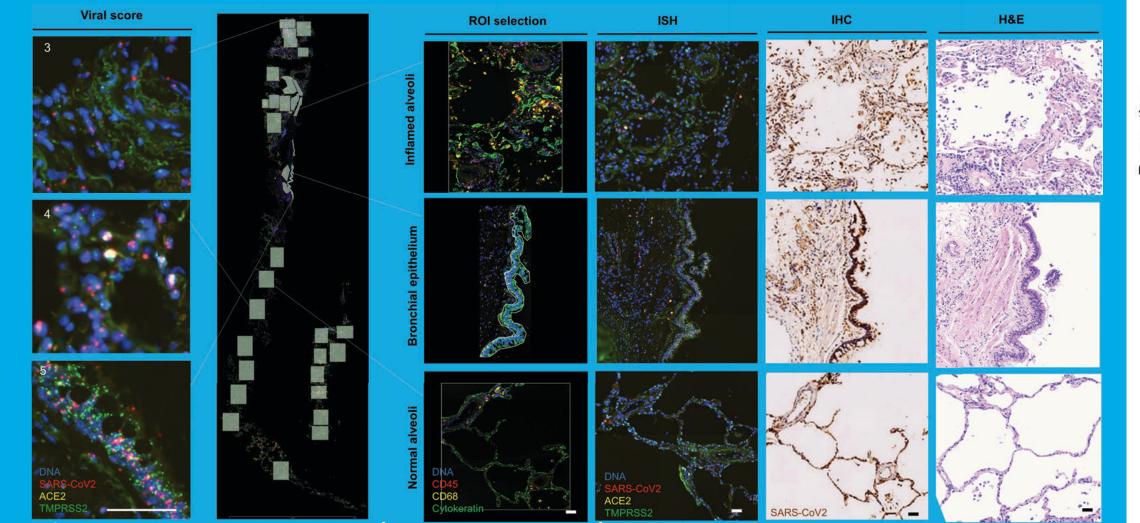
Yered Pita-Juarez<sup>1,2,3</sup>, Liuliu Pan<sup>4</sup>, Jonathan Hecht<sup>1,2</sup>, Tyler Hether<sup>4</sup>, Linus Tsai<sup>2,5</sup>, Yury Popov<sup>2,5</sup>, Jason Reeves<sup>4</sup>, Pourya Naderi<sup>1,2</sup>, Sebastian Niezen<sup>2,5</sup>, Stefan Riedel<sup>1,2</sup>, Eric Miller<sup>4</sup>, Winston Hide<sup>1,2</sup>, Sarah Warren<sup>4</sup>, Gyongyi Szabo<sup>2,3,5</sup>, Joseph Beechem<sup>4</sup>, Z. Gordon Jiang<sup>2,5</sup>, Ioannis S. Vlachos<sup>1,2,3</sup>

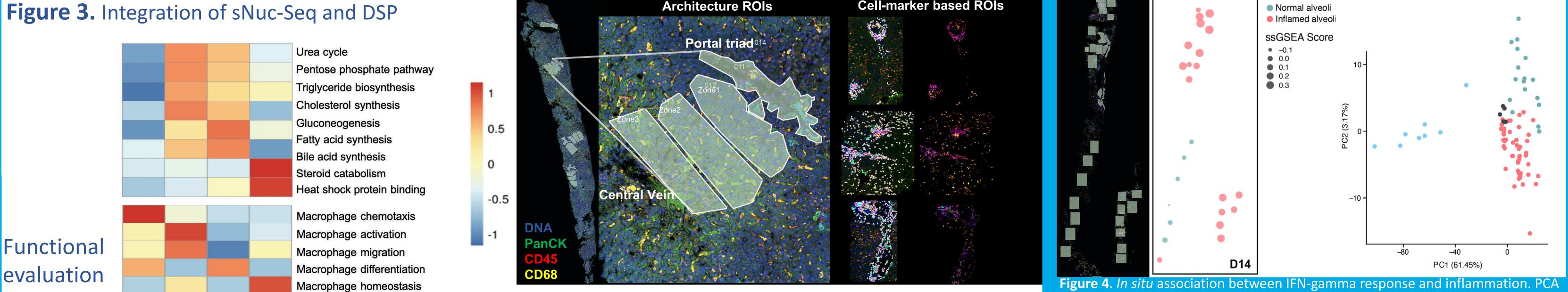
ID	Age/Sex	Hospital stay (d)	Comorbidities	SARS-CoV2: lung (Cp/ug RNA)
013	80+M	10	ESRD on dialysis, HTN	6.0x 10 <sup>7</sup>
14	80+F	16	HTN, DM2, TIA, dementia	2.1 x 10 <sup>4</sup>
<b>)15</b>	50-60F	40	HTN, DM2, obesity, CKD	10
16	70-80F	15	ESRD on dialysis, HTN	1.7x10 <sup>4</sup>
)17	70-80M	31	Myasthenia gravis, HTN	8,435

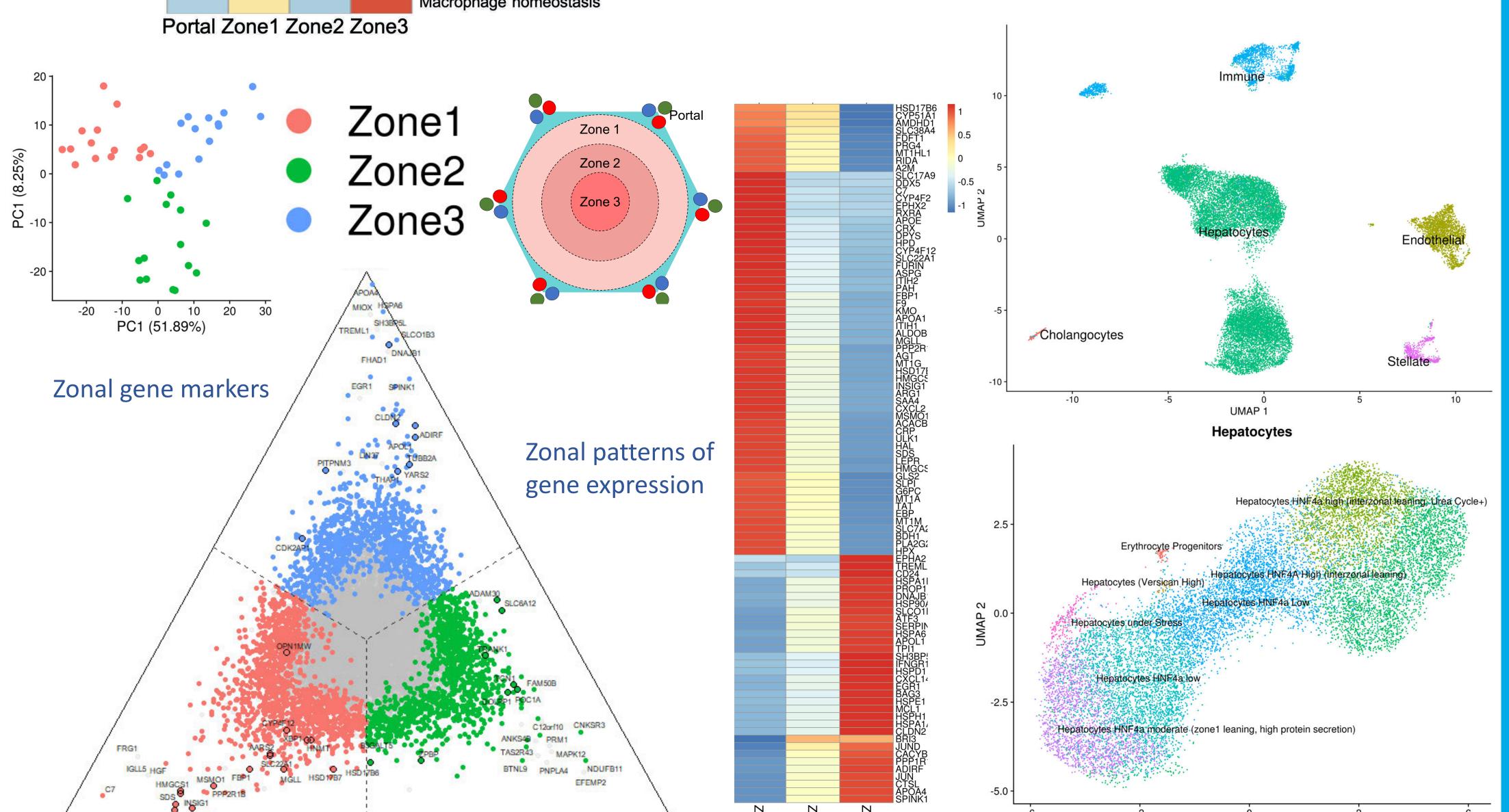
Functional

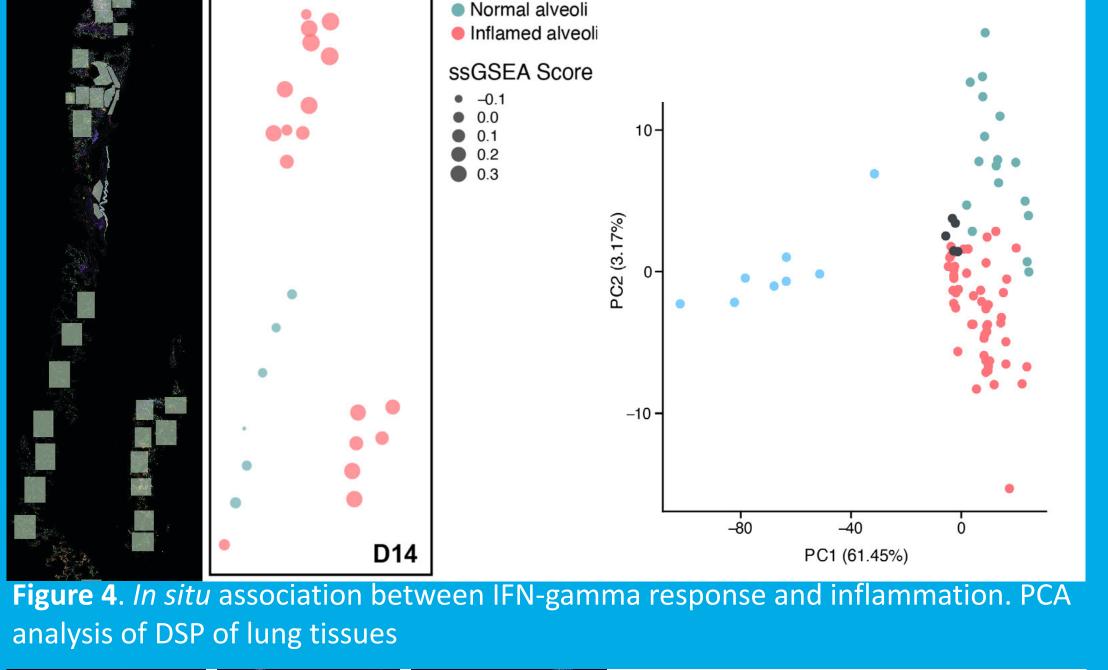
evaluation

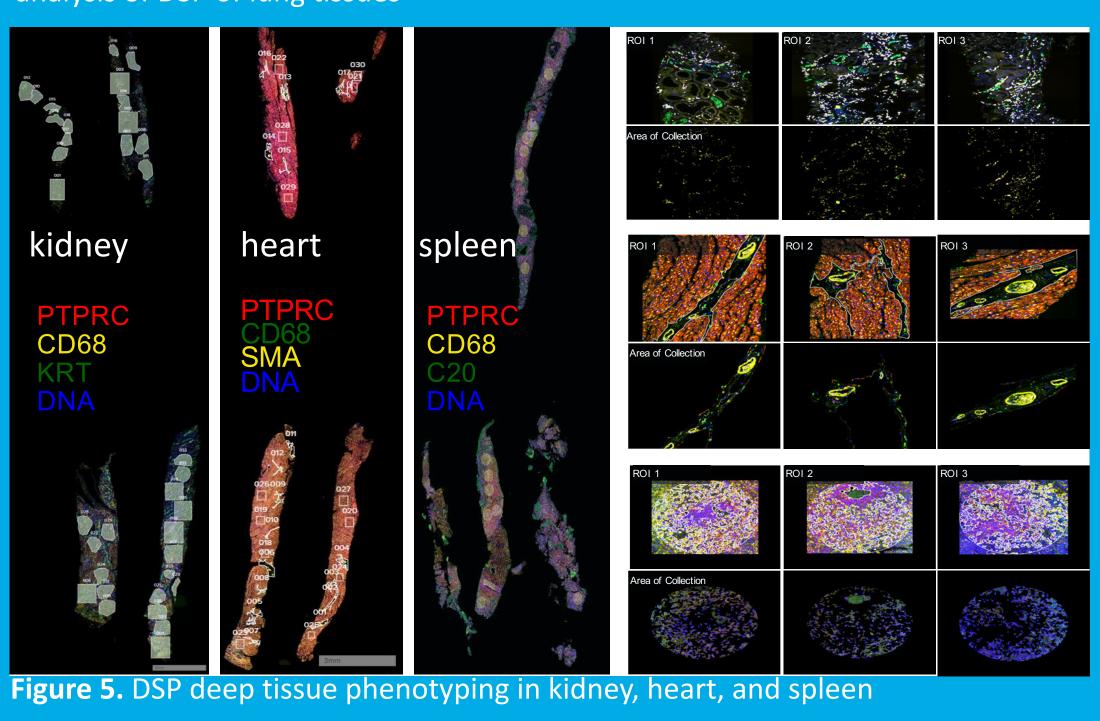


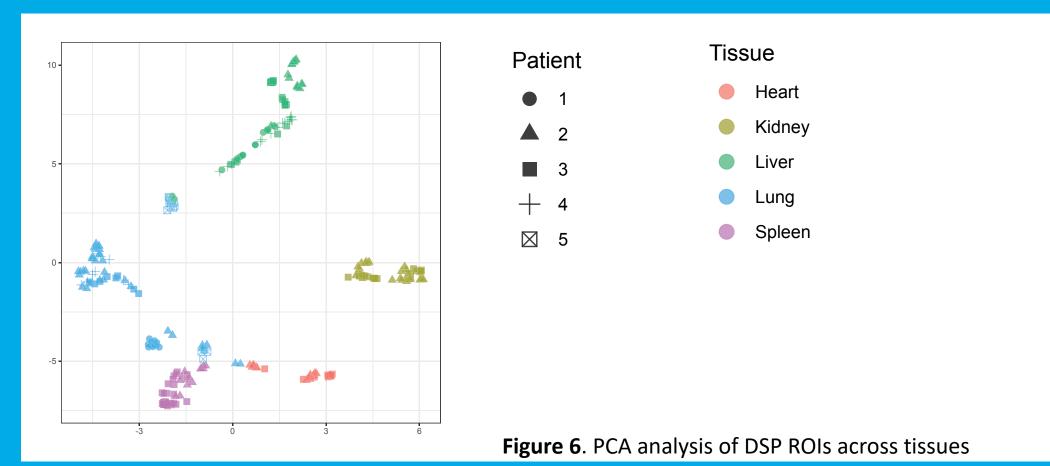








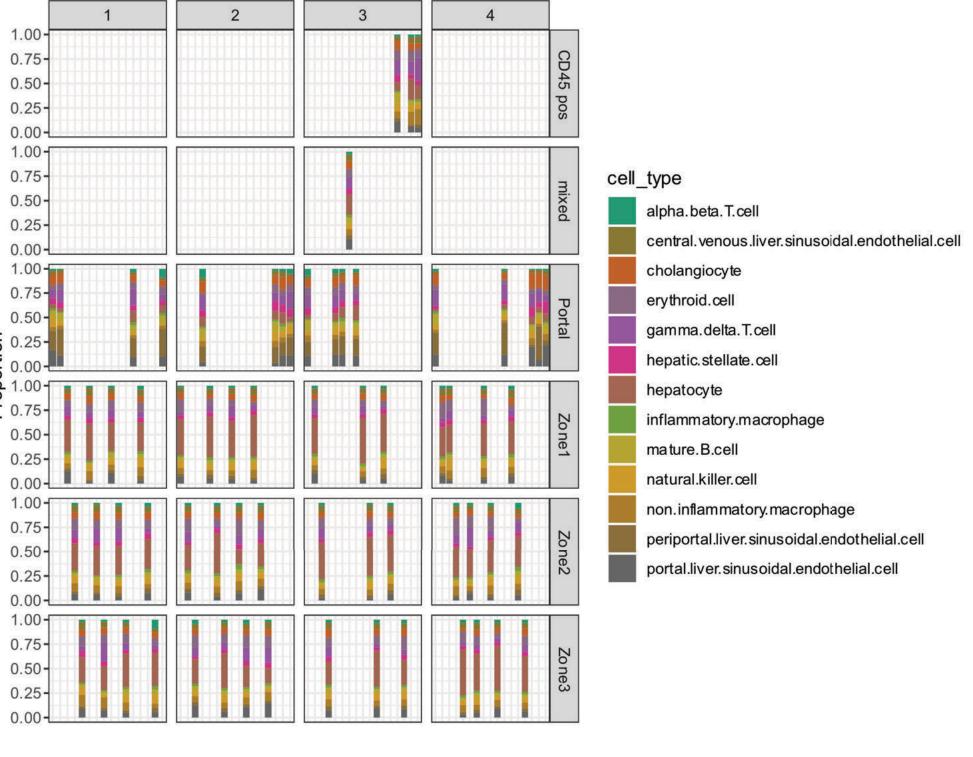




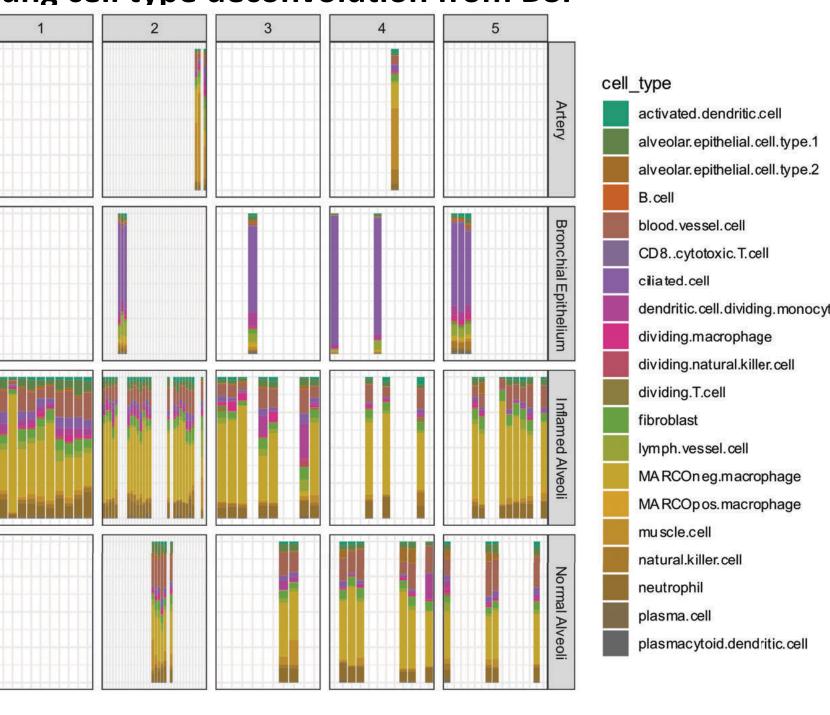
### NanoString products are For Research Use Only and not for use in diagnostic procedures

## Supplementary

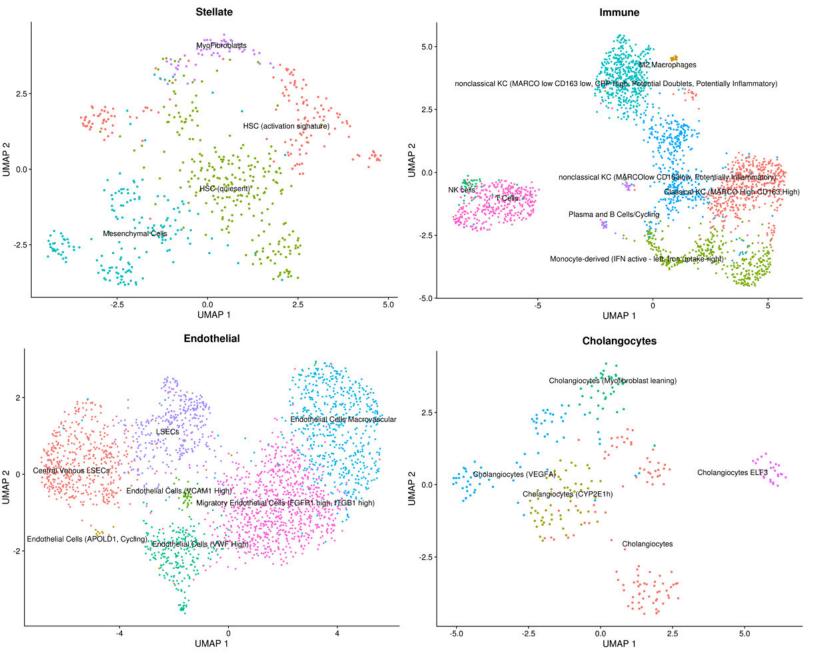




## Lung cell type deconvolution from DSP



## Non-Parenchymal Liver Cells (sNuc-Seq)



## Acknowledgements

#### We are deeply grateful to all donors and their families.

We would like to thank Dimitra Pouli who supported the creation of tissue annotation

G.S. acknowledges support from the NIH R01AA0207440 and U01AA026933 research



