nanoString

TruCulture®-NanoString® Workflow

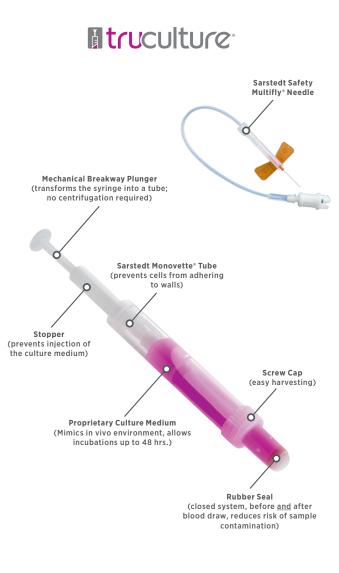
Sample Extraction and Preparation

- 1. After incubation, separate cells from supernatant using Seraplas valve and plunger
- 2. Remove supernatant
- 3. Remove Seraplas valve
- Remove cell layer (~1mL) using a pipette to fresh 15 mL conical tube containing 3 mL RNAlater
- 5. Once resuspended in RNAlater, cells can be stored or immediately processed by RNA extraction

RNA Extractions

- 6. Gently vortex sample tube for 5 seconds to mix cells
- **7.** Remove appropriate volume of cell suspension for RNA extraction method being used
 - RNA is extracted using the Ambion RiboPure RNA kit with a recommended column capacity of 500 μL whole blood
 - 2 mL cell suspension (cell equivalent of 500 µL whole blood) is aliquoted to a fresh tube for the Ambion protocol
- 8. Add 2 mL PBS and mix thoroughly
- 9. Centrifuge at 2000g for 10 minutes (until cells are pelleted) and remove supernatant
- 10. Continue with step B1.b of the Ambion RiboPure protocol, and continue with protocol to completion

Note: The Ambion protocol recommends 100 µL elution volume. RNA concentration may be increased by reducing elution volume if desired



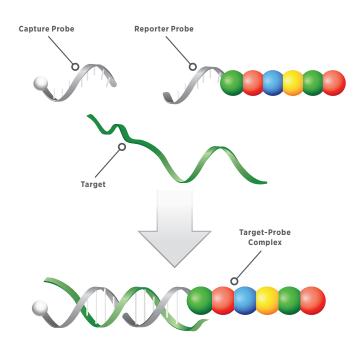
(cont.)

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TruCulture®-NanoString® Workflow (cont)

RNA QC and NanoString Assays

- **11.** Measure RNA concentration by A260 UV absorbance (e.g., Nanodrop) or alternatively by fluorescent dye method (e.g., Qubit)
 - Normalize RNA to a single concentration (e.g., 20 ng/µL for a 100ng total concentration) by dilution in appropriate volume of molecular biology grade water.
- 12. Set up nCounter[®] hybridizations following NanoString MAN-10023 and continue with downstream processing and data collection as appropriate for your nCounter system.
 - Recommended minimum RNA input is 25 ng.



For more information, visit nanostring.com/truculture

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