



Unlocking Predictive Potential for Response to Immunotherapy

Digital Spatial Profiling Technology

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Unlocking Predictive Potential for Response to Immunotherapy: Digital Spatial Profiling Technology

Introduction

Stage III melanoma is characterized by the spread of the cancer cell from the skin into the lymph nodes. Surgery is the first line of treatment to remove the tumor(s), cancerous lymph nodes, and healthy tissue adjacent to the tumors. Often adjuvant therapy is administered post-surgery. This can include weeks of radiation, chemotherapy, targeted drug therapy, or immunotherapy. These therapies can have mixed results and side effects that vary by patient. Researchers are actively investigating these differences in outcome to identify combinations of biomarkers that may predict a patient's response to treatment. These expression signatures may help guide the physician to administer more effective treatments in a deliberate, evidence-based manner.

The challenge today is in identifying these combinations of biomarkers at play in the tumor microenvironment. Quantitating the levels of proteins and RNAs in a tumor sample often requires destroying the tissue and sacrificing spatial information. Alternatively, while fluorescence and brightfield imaging provide

a visual map of expression they are limited by the number of fluorophores that can be captured in one experiment. While it is possible to perform multiple rounds of immunostaining and imaging on the same sample, the sample will degrade over time and errors in image registration can lead to misinterpretation of results.

The GeoMx™ Digital Spatial Profiler (DSP) is a new platform that leverages the nCounter® barcoding technology to both spatially resolve and digitally quantify protein and mRNA expression. The assay uses either antibody or RNA probes coupled to photocleavable oligonucleotide tags. After binding of oligo-conjugated probes and up to four morphology markers (fluorescent probes) to slide-mounted formalin fixed paraffin-embedded (FFPE) tissue sections, the oligonucleotide tags are released from selected regions of the tissue by UV exposure. Released tags are quantitated in a standard nCounter assay, and counts are mapped back to tissue location, yielding a spatially-resolved digital profile of analyte abundance (**FIGURE 1**).

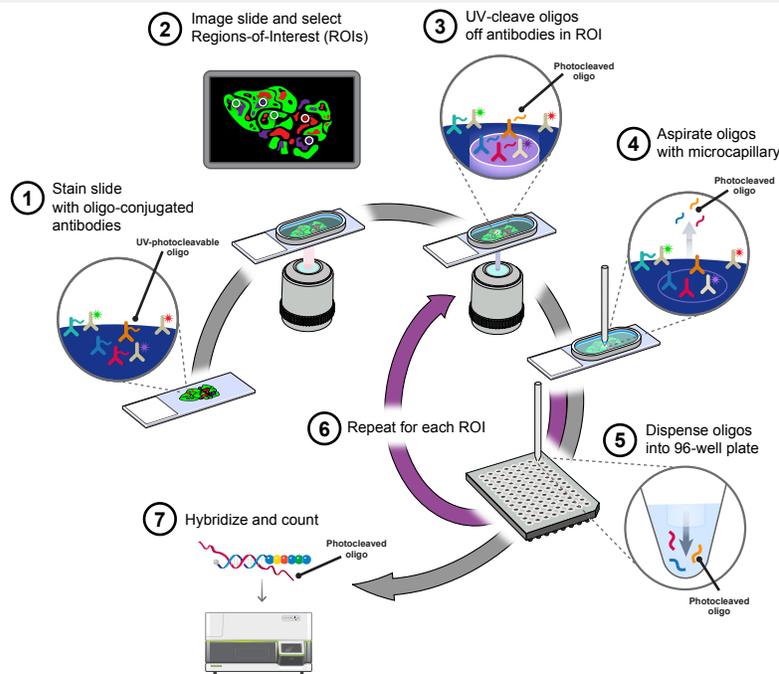


FIGURE 1: GeoMx DSP Overview: Samples are stained with oligo-conjugated antibodies, imaged and ROIs selected. Oligos are then cleaved from the targeted ROIs, collected, and analyzed.

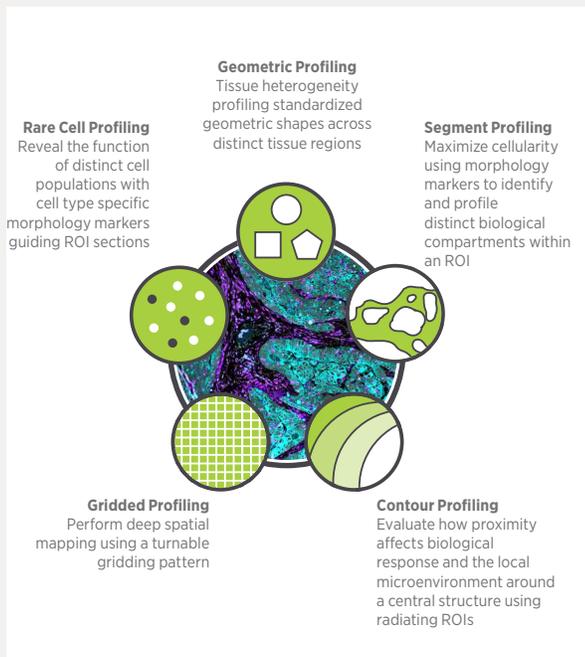


FIGURE 2: Five profiling modalities of GeoMx DSP

GeoMx DSP Profiling Modalities

Five main GeoMx DSP profiling modalities have been identified to define regions of interest (ROI): Geometric, Segment, Contour, Gridded, and Rare Cell (**FIGURE 2**). Geometric profiling uses geometric shapes to quantify expression within the chosen boundaries. The same shape can be reused, ensuring that the specific area (in pixels) is the same between ROIs. Segment profiling recognizes the differences between high and low signals from morphology markers (fluorescent targets) to identify and profile distinct biological areas within a ROI, for instance CD45-positive versus S100B-positive tissue (**FIGURE 3**). Contour profiling can illustrate how proximity affects biological response by examining the local microenvironment around a central structure using radiating ROI. Central structures can be compact, such as clusters of immune cells, or complex, like a neuron or blood vessel. Gridded profiling creates a tunable gridding pattern that is overlaid on the image to drive deep spatial mapping of a sample. Finally, Rare Cell identifies distinct cell populations based on cell type specific morphology markers, shining a spotlight on rare events in an otherwise crowded field.

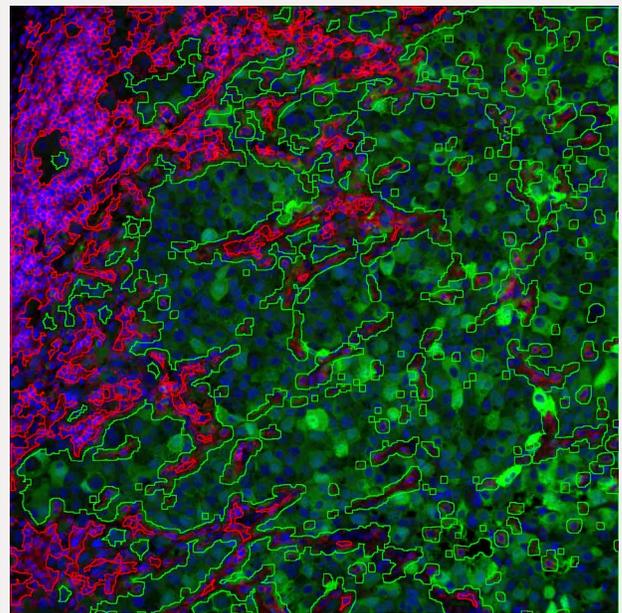
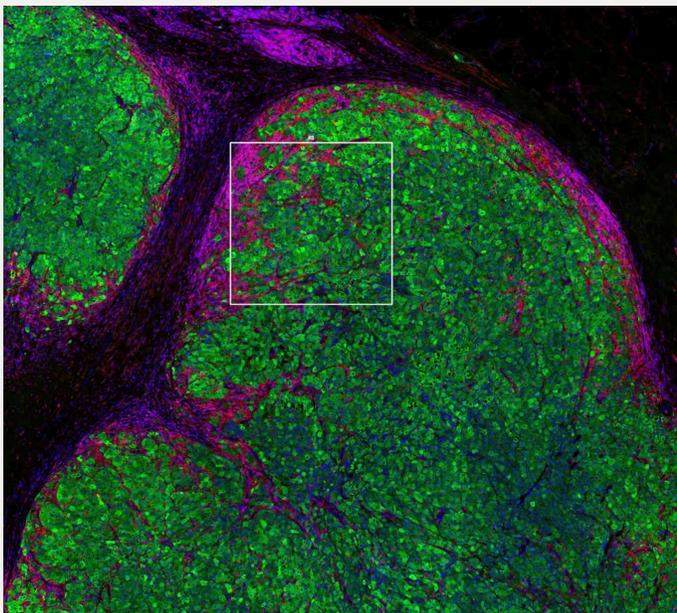
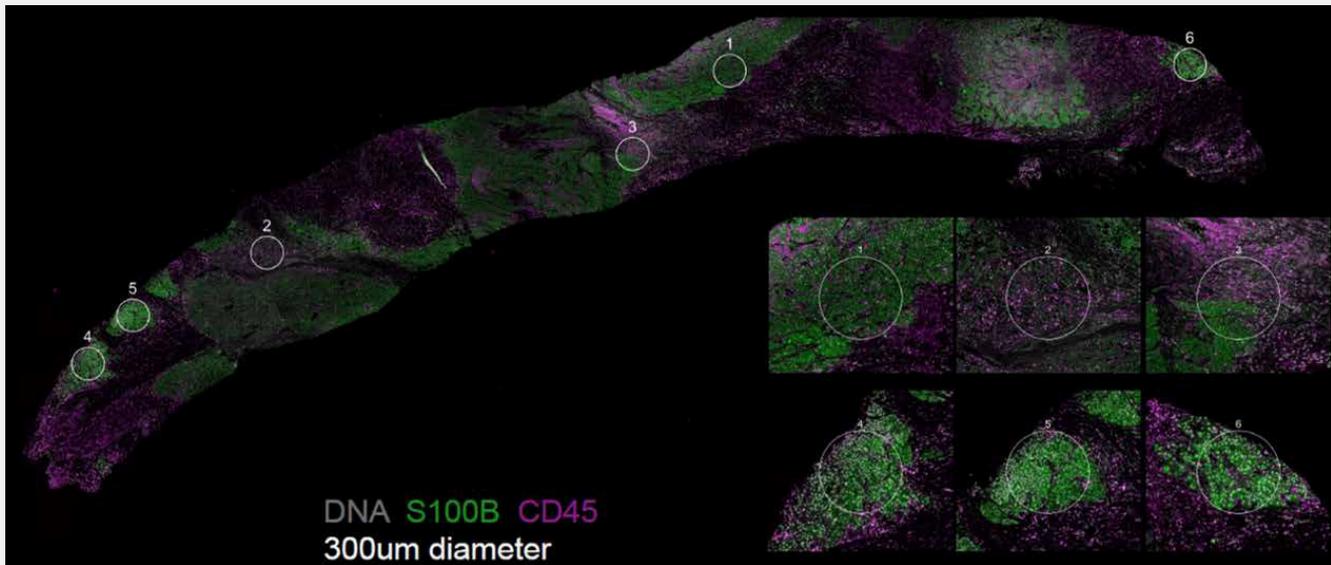


FIGURE 3:

- A. Tumor sample stained with S100B (tumor cells) and CD45 (immune cells). Segments are generated based on S100B and CD45 cellular morphology.
- B. Enlarged view of segmentation: Green= S100B-positive tumor cells, red= CD45-positive immune cells, and blue = DNA. Each segment is collected and quantified separately within the ROI.

A



B

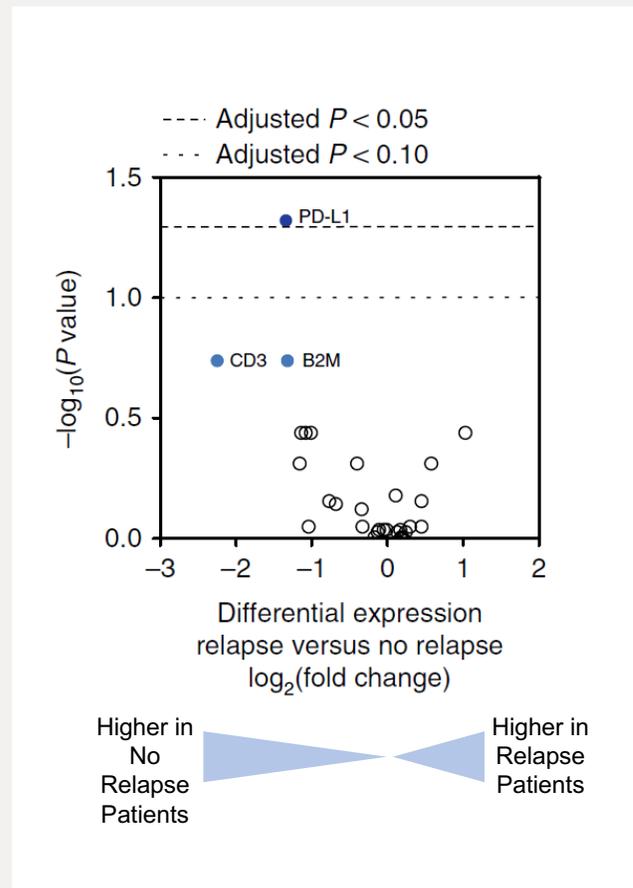


FIGURE 5:

- A. Geometric ROI selection strategy
- B. Volcano plot measuring differential expression of proteins between patients with melanoma that relapsed or did not relapse after neoadjuvant therapy analyzed by GeoMx DSP. Note the increased levels of $\beta 2M$, CD3, and PD-L1 in patients without relapse. None of the other proteins are shown in the figure and CD3 is associated with the adaptive immune response (figure reproduced from Nature Medicine).

Blank, CU, et al. Neoadjuvant versus Adjuvant Ipilimumab Plus Nivolumab in Macroscopic Stage III Melanoma. Nat Med. 2018; 24(11): 1655-1661.

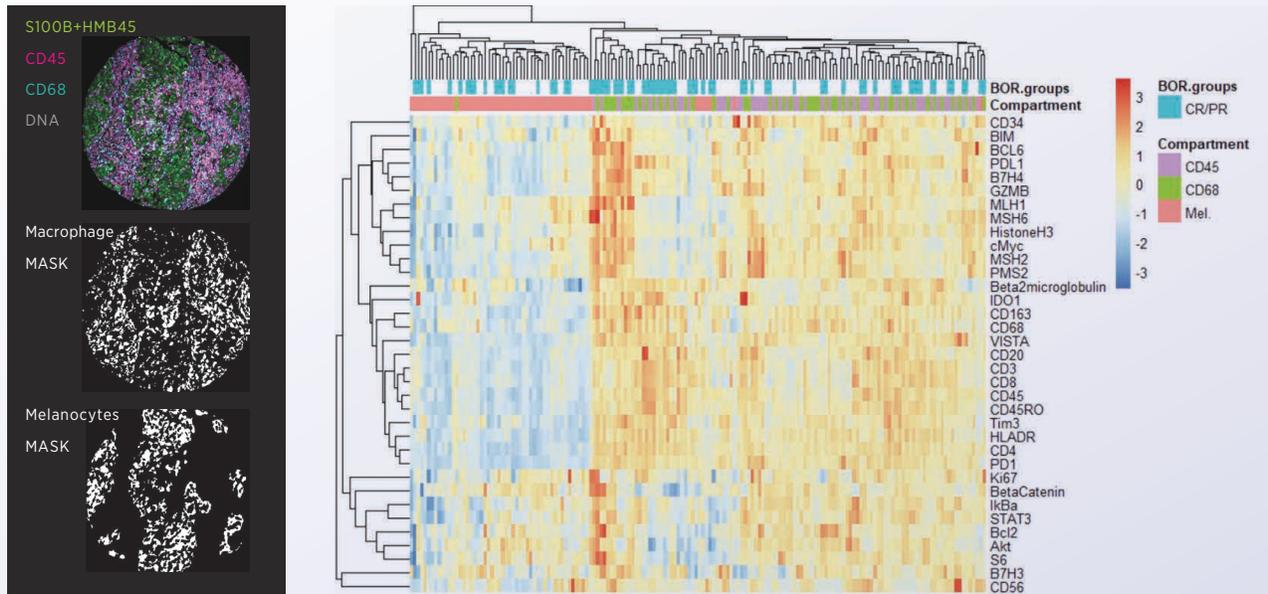
Geometric Profiling. CD45 staining was also used to establish three ROIs with high immune infiltrate and three ROIs with low immune infiltrate. The group quantified levels of CD3, β -2 microglobulin, and PD-L1 protein with GeoMx DSP and also stratified IFN- γ RNA levels as low, intermediate, and high. Patients with decreased levels of CD3, β -2 microglobulin, and PD-L1 and low levels of IFN- γ RNA relapsed. Patients with intermediate to high levels of IFN- γ RNA did not relapse (at time of publication), indicating that this biosignature has the potential to be used to predict the patient's response to treatment (**FIGURE 5**).

Recent work from David Rimm and colleagues at Yale University looked at specific cell types within the tumor microenvironment to identify prognostic biomarkers. Compartments were elucidated with the Rare Cell Profiling using serial masks, focusing on macrophage (CD68+), melanocyte (S100B+), and non-macrophage immune cells (CD45+CD68-). The team was particularly interested in differentiating between the tumor and the stromal areas. CD3, CD8, β -2 microglobulin, PD-L1, and HLA-DR all demonstrated cell type-specific predictive power both in overall survival rates and progression free survival. PD-L1 showed strongest association with overall survival in the macrophage compartment (**FIGURE 6**). β -2 microglobulin in the immune, non-macrophage compartment was associated with both overall survival and progression free survival. With this information, the group has identified compartment specific markers associated with potential prognostic biomarkers of survival.

Conclusion

Using GeoMx DSP technology, potential predictive biomarkers have been identified for clinical responses to both ipilimumab and nivolumab. Whether used as neoadjuvant or adjuvant therapies, there is an inherent toxicity associated with these checkpoint inhibitors. Side effects of nivolumab and ipilimumab can range depending on the patient and can be severe enough to discontinue use. While at this time the side effects one might experience cannot necessarily be predicted, we are gaining a better understanding of whether a therapy will be beneficial prior to beginning treatment.

Moving beyond ipilimumab and nivolumab, there are many other single agent and combination immunotherapies that still need to be characterized for predictive biomarkers. GeoMx DSP has shown results that strongly track with established immunohistochemistry and pathology methods and offers the potential opportunity to profile and establish more biomarkers in a single experiment than these techniques. Moreover, its utility to perform high-plex analysis of RNA and protein for discovery of prognostic and predictive markers holds great potential for application and advancement in clinical practice and medical technology.



High PD-L1 expression in macrophages is associated with prolonged survival

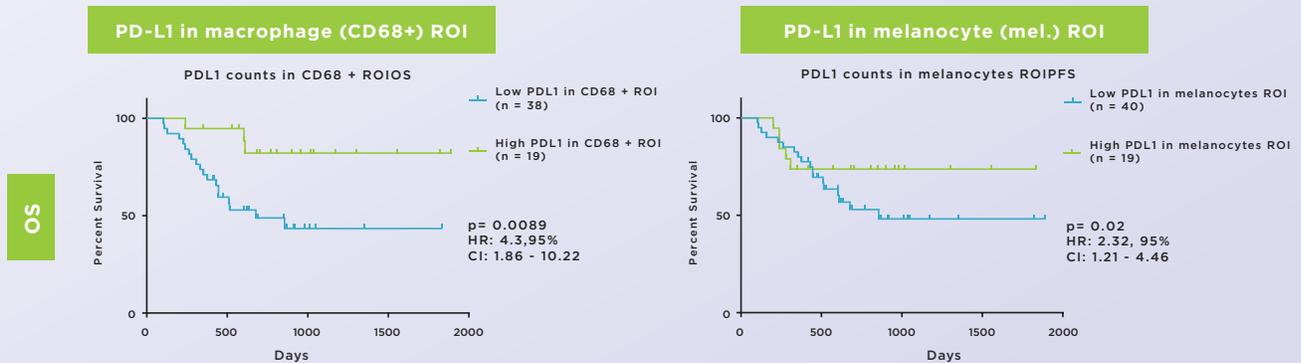


FIGURE 6: PD-L1 expression specifically in macrophages potentially predicts outcome. 44 proteins were profiled across 3 unique compartments from 59 immuno-therapy treated melanoma patients using rare cell profiling. Five compartment specific biomarkers were identified with PD-L1 expression in macrophages showing the strongest predictive power.

- 1 Weber, J, *et al.* Adjuvant Nivolumab versus Ipilimumab in Resected Stage III or IV Melanoma. *N Engl J Med.* 2017; 377(19): 1824-1835.
- 2 Eggermont, AM, *et al.* Prolonged Survival in Stage III Melanoma with Ipilimumab Adjuvant Therapy. *N Engl J Med.* 2016; 375(19): 1845-1855.
- 3 Wolchok, JD, *et al.* Overall Survival with Combined Nivolumab and Ipilimumab in Advanced Melanoma. *N Engl J Med.* 2017; 377(14): 1345-1356.
- 4 Amaria, RN *et al.* Neoadjuvant Immune Checkpoint Blockade in High-risk Resectable Melanoma. *Nat Med.* 2018; 24(11): 1649-1654.
- 5 Blank, CU, *et al.* Neoadjuvant versus Adjuvant Ipilimumab Plus Nivolumab in Macroscopic Stage III Melanoma. *Nat Med.* 2018; 24(11): 1655-1661.

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